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Combination Therapy with Vitamin D and Telmisartan to Suppress the

Review Article

Progression of Liver Fibrosis.

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ABSTRACT

Hepatic fibrosis is an improper wound repair response associated with various kinds of chronic liver injuries. It can be detected by over-deposition of diffuse ECM (Extracellular Matrix) and abnormal overgrowth of connective tissue, and it can progress to cirrhosis or hepatocellular carcinoma. Chronic liver diseases, which include liver fibrosis, have resulted in significant morbidity and mortality worldwide, and this trend is likely to continue. Although early liver fibrosis has been reported to be reversible, the exact mechanism of reversal is unidentified, and there is a lack of effective therapy for liver fibrosis. Vitamin D is essential for maintaining the metabolism of bone and calcium balance. Unexpectedly new evidence indicates that vitamin D has a protective function toward hepatic fibrosis. Telmisartan has been shown to have a positive effect of vitamin D and telmisartan on the progression of hepatic fibrosis and updating their present impact on fibrosis. Telmisartan and vitamin D reduce fibrosis in hepatic stellate cells by inhibiting the production of profibrogenic genes.

Keywords: Liver fibrosis, Vitamin D, Vitamin D receptor, Telmisartan

INTRODUCTION

The liver is the most essential organ in humans, it is necessary for many processes like the metabolism of energy, toxic substance elimination, and control of the immune system. Because of these intensive functions, the liver is susceptible to a variety of diseases, including viral infections, autoimmune diseases, malnutrition, and alcoholism. Continuous damage to the liver has been shown to result in hepatic fibrosis. ^[1, 2].

Liver Diseases

The liver, as the primary organ for detoxification and metabolization, is vulnerable to a variety of illnesses. Liver disease rates have progressively risen throughout the years. According to WHO, chronic diseases cause approximately 46% of the world's diseases and 59% of mortality, with nearly 35 million people dying worldwide ^[3]. Several major diseases of the liver can lead to inflammation. This inflammation can lead to scarring or cirrhosis.

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Diseases can be classified as either acute or chronic [4]. There are numerous types of liver disease, which can be a consequence of a virus, medication or chemical destruction, obesity, diabetes, or an immune system attack. They are classified as hepatic steatosis, jaundice, hepatitis, fibrosis, cirrhosis, cholestasis, and carcinoma, as indicated in *Figure 1*^[5].

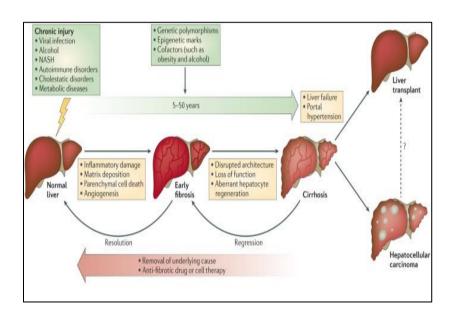


Figure 1. Chronic hepatic diseases development ^[6]

Liver fibrosis

Hepatic fibrosis is a serious disorder that may result in cirrhosis and liver cancer. There are several types of chronic liver damage, like viral infections, cholestatic diseases, consumption of alcohol, nonalcoholic steatohepatitis, and non-alcoholic fatty liver disease, which may result in inflammation of the liver and abnormal wound healing process, resulting in fibrosis ^[7]. Hepatic fibrosis can be recognized by excessive accumulation of extracellular matrix (ECM) and formation of fibrous scar. degradation of liver structure by fibrous scars and the loss of hepatocytes could inhibit the biological functions of the liver, leading to liver failure ^[8]. The evaluation of fibrosis allows significant information and is highly helpful for the diagnosis, in addition to the treatment strategy and evaluating the natural history or progression during therapy.^[9].

The pathological process of Liver Fibrosis

Alcoholism, metabolic problems, viral infection, obesity, steatosis, and cholestasis are all risk factors for liver fibrosis, with abuse of alcohol being the most common. Throughout the metabolism of alcohol, acetaldehyde, and ROS (reactive oxygen species) are formed. Acetaldehyde stimulates TGF- β synthesis and type I collagen production in HSCs, leading to liver fibrosis ^[10, 11]. The pathogenic liver fibrosis process mostly involves the accumulation of collagen and proteins of ECM in fibroblasts that trigger wound repair responses ^[12]. Myofibroblast stimulation and proliferation are the most common causes of fibrous collagen and the deposition of ECM in the injured liver. During HSC activation, Myofibroblasts are produced and released from portal fibroblasts, bone marrow-derived fibroblasts, mesenchymal cells, and liver parenchyma-derived myofibroblasts by epithelial-mesenchymal

transition (EMT), resulting in a major portion of the liver fibrosis ^[13].

Pathogenesis of liver fibrosis determines which different types of myofibroblasts develop. Previous research has shown that the primary source of myofibroblasts in the hepatic fibrosis model induced by CCl₄ is HSCs. In the cholestatic liver, portal fibroblasts generate myofibroblasts, while in chronic harm, bone marrow-derived cells contribute to the total fiber-derived differentiation ^[14-16]. Progression of liver fibrosis is mainly controlled by different cells and cytokines, and HSC activation is considered the main link of hepatic fibrosis ^[17, 18].

HSCs are located in the subcutaneous space surrounding the liver's sinuses, among hepatocytes and blood sinus endothelial cells. In normal physiological processes, HSCs are in dormancy and store fat and fat-soluble vitamins. In liver damage, HSCs are triggered by inflammatory mediators, which then divide into myofibroblasts. After that, HSCs release proteins of ECM and matrix metalloproteinases, which result in remodeling of liver tissue. progression of hepatic fibrosis can be seen in *Figure 2*^[19]

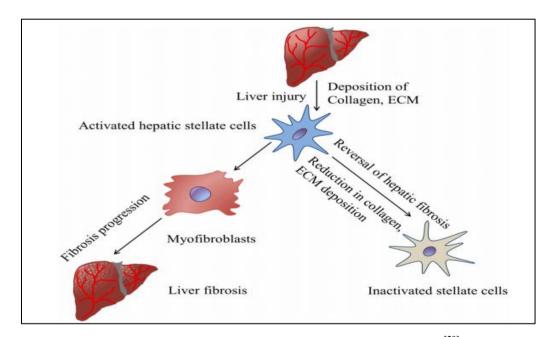


Figure 2. Development and reversal of hepatic fibrosis.^[20]

Diagnosis of hepatic fibrosis

Precise diagnosis of the degree of hepatic fibrosis is required for medical therapy to assess the prognosis and treatment decisions among patients ^[21]. Despite the development of possible diagnostic procedures over the last 50 years, biopsy is regarded as the gold-standard technique for assessing fibrosis and provides helpful details about diagnosis and other harmful processes such as necrosis, inflammation, and steatosis ^[22]. The Ishak score, Metavir score, and Desmet/Scheuer staging system are three commonly applied for assessing histological fibrosis ^[23].

The scoring system depends on the progression of periportal fibrosis, after septal fibrosis, followed by nodule formation ^[24]. One of the liver biopsy limitations is its high invasiveness. Furthermore, low quality of the sample and tissue size result in non-reproducible biopsy, and it is dependent entirely on the pathologist's knowledge, resulting in

interobserver variations. Due to the drawbacks of liver biopsy, a noninvasive method for diagnosing liver fibrosis was developed. A suitable biomarker should be organ-specific, sensitive to detecting active damage, easily accessible in peripheral tissue, and inexpensive ^[25].

The advantages of biomarkers in comparison to liver biopsy are that they can be estimated in serum using a minimally invasive method. Additional benefits include ease of use, inter-laboratory reproducibility, and widespread availability. Serum biomarkers of hepatic fibrosis can be categorized into 2 different groups: direct biomarkers, which indicate turnover of ECM, and indirect biomarkers, which are released into the blood and indicate changes in hepatic function ^[26]. Indirect markers indicate changes in liver function. These markers may assist with liver disease diagnosis, severity evaluation, therapy monitoring, and prognosis assessment. Measuring enzyme activity, including levels of aminotransferases, alkaline phosphatase (ALP), and γ -glutamyl transferase (γ GT), and also estimating bilirubin and albumin in the blood ^[27, 28].

Direct markers play a direct role in the accumulation of ECM made by HSC and other hepatic cells. Levels of these markers in serum are increased in the progression of fibrosis and tend to decrease during therapy ^[9]. these markers Evaluation may be beneficial in determining efficient therapy. Direct markers can be categorized by their molecular structure ^[29].

Treatment of hepatic fibrosis

The present therapies for liver fibrosis attempt to eliminate related damage factors by inhibiting hepatic stellate cells (HSCs) activation, promoting ECM destruction, and preventing inflammatory reactions ^[30, 31]. Currently, there are no approaches to treating liver fibrosis or early cirrhosis in clinics other than conventional drug therapy. Conventional therapies have some drawbacks for tissues and organs, including toxic effects and side effects, in addition to Drug specificity is low, this prevents

being therapeutic drugs from effectively concentrated in the liver. As a consequence, their efficacy for treatment is not desirable^[32]. As a result, the article's primary goal was to assess vitamin D telmisartan's beneficial effects on and the progression of hepatic fibrosis, focusing on their critical mechanisms controlling liver fibrosis. Furthermore, we sought to summarize the current state of their treatment in hepatology. Particularly, approaches for both early detection and therapy of liver fibrosis.

Role of Vitamin D in liver fibrosis

Vitamin D is a steroidal hormone that plays a major role in maintaining calcium and bone balance. ^[33]. Besides its traditional effect on the health of bones, it has a biological effect on many types of cells, resulting in control of proliferation and differentiation of cells ^[34]. Vitamin D is available in 2 equivalent types: vitamin D₂ and vitamin D₃, each of which can be obtained through diet.

Vitamin D₂, or ergocalciferol, is produced in certain kinds of plants but primarily in fungal organisms through UVB (ultraviolet B) activity on ergosterol. In humans, the skin produces the greatest amount of vitamin D3 through UVB rays from In epidermis, 7sunlight. the lower dehydrocholesterol (DHC) is converted to previtamin D₃. A thermal-dependent isomerization process converts pre-vitamin D₃ into vitamin D³ (cholecalciferol)^[35]. Telipophilic cholecalciferol is non-bioactive. It must also go through two sequential hydroxylation in the liver and kidney to become an intermediate metabolite and then reach its final active form. In the blood, cholecalciferol is primarily linked to vitamin D-binding protein (DBP) or albumin before reaching the liver for 25hydroxylation. 25-OHD₃ is commonly used for determining systemic vitamin D levels. In the kidney's proximal tubule, 25(OH)D-1a-hydroxylase enzyme, or CYP27B1, turns 25(OH)D3 to its active type [36].

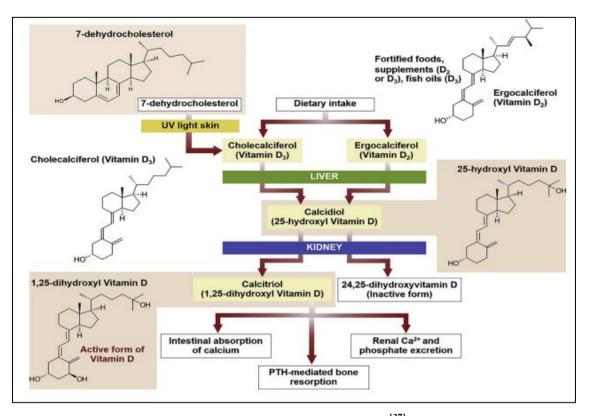


Figure 3. Vitamin D Biosynthesis^[37].

Vitamin D effect on liver diseases

Vitamin D helps to reduce the risk of chronic diseases such as diabetes, cancer, and other heart disease, autoimmune, and infectious diseases. the effect is due to the synthesis of 1α , 25(OH)2D, which autocrine and paracrine processes has in proliferation and differentiation and cell death [36, 38]. Recently, vitamin D plays an essential part in maintaining the fibrotic process, inhibiting collagen synthesis in stromal HSCs. Additionally, data are building up to support the theory that vitamin Dmediated VDR activity could be involved in the reduction of fibrosis. For instance, current research by Ding et al. ^[37] VDR enhances Unexpected fibrosis of the liver.

The vitamin D/VDR direction inhibits TGF- β -induced fibrosis of the liver in HSCs by attaching with regulatory loci of pro-fibrotic genes which decrease SMAD-3 recruitment. This VDR/SMAD genomic feedback process regulates fibrosis. In Abramovitch et al. [39] research, They discovered the anti-fibrotic activity of vitamin D in HSCs by

responding to VDR, implying a possible physiological effect on VDR-mediated fibrosis. But Neeman et al. [40] found that farnesylthiosalicylic acid and vitamin D had a beneficial impact on HSCs by the Ras-guanosine-5'-triphosphate (GTP) and phospho-extracellular signal-regulated kinase (pERK) signaling pathways. Artaza and Norris showed that vitamin D reduces TGF- β by blocking many pro-fibrotic proteins.

Vitamin D inhibits the production of collagen I and III was inhibited by vitamin D while increasing the production of MMP-8 [41]. A metalloproteinase plays an important role in restricting extracellular matrix destruction that is linked to the development fibrogenesis. Activating TGF-β/SMAD-3 of promotes fibrosis by transforming HSCs and matricellular protein secreting а involving connective tissue growth factor at the destruction of ECM [42]. in rats, CTGF synthesis was significantly raised; but after vitamin D medication, its levels significantly dropped ^[43]. the results demonstrate that there is an important role of vitamin D-mediated

CTGF expression in the progression of fibrogenesis [44]

Results from in vitro and in vivo research further indicate vitamin D's protective role in fibrogenesis. Recently, Beilfuss et al. ^[45]. discovered that vitamin

D/VDR inhibits TGF- β -induced pro-fibrogenic gene expression in HSCs. Also, there is a correlation between VDR and fibrosis among nonalcoholic fatty liver disease patients as shown in *Figure 4*.

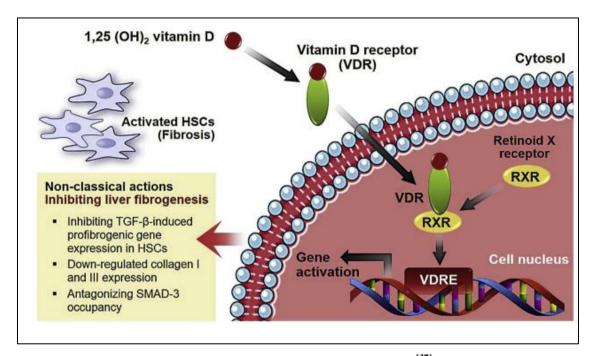


Figure 4. Vitamin D and hepatic fibrosis [37].

These consequences have an important part in chronic liver diseases. Deficiency of Vitamin D is particularly known among individuals who have chronic liver disease. 93% of these people have vitamin deficiency ^[46, 47]. Even those with mild liver disease are affected. Patients suffering from liver diseases. Up to 93% of these patients have some level of vitamin deficiency. Multiple investigations demonstrated that tiny amounts of 25(OH)D increase the chance of mortality, involving heart disease ^[48].

Role of Telmisartan in liver fibrosis

Telmisartan is a particular angiotensin receptor blocker (ARB) that regulates PPAR-γ activity, enhancing insulin sensitivity and reducing hepatic fat building up ^[49], Furthermore, blocking the angiotensin II receptor prevents activation of HSC, so it reduces fibrosis ^[50, 51]. Telmisartan decreases hepatic damage caused by type 1 diabetes mellitus ^[52], Combined with propranolol, it decreases signals of fibrosis like hydroxyproline, bile duct development, procollagen- α 1, endothelin-1, and metalloproteinases in a PSC-like mouse model ^[53]. It additionally prevents fibrosis in the rat bile duct ligation model ^[54].

Telmisartan reduces liver inflammation and fibrosis in rats who nourished a fatty diet and provided a small dosage of streptozotocin (STZ) just two days after birth ^[55]. It acts as an angiotensin receptor blocker that treats hypertension. Telmisartan (Micardis®; Boehringer Ingelheim, Ingelheim, Germany) is a nonpeptide ARB with orally active properties that are linked to AT1 receptors, preventing angiotensin II's biological effect. It has a bis-benzimidazole structure, as illustrated in *Figure 5*.



138

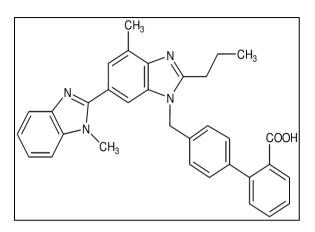


Figure 5. Chemical Structure Of Telmisartan^[56].

Additionally, it is helpful for hypertension, improving the sensitivity of insulin in diabetes Type 2, and lowering triglycerides. Telmisartan can be used to treat NASH with metabolic syndrome, as the majority of NAFLD patients have metabolic syndrome features ^[57]. ARBs have been widely recognized as a method of blocking the contraction of blood vessels and inhibiting the growth of cells and fibrogenesis which is controlled via the angiotensin II type 1 receptor (AT1-R) ^[58]. AT1-R knockout mice exhibited lower levels of hepatic TGF-β1 and pro-inflammatory cytokines in comparison to WT mice. Moreover, Bataller et al ^[59]

Bataller et al[61] found that elevated systemic Angiotensin (Ang) II levels cause fibrogenesis and promote inflammation. Telmisartan has the strongest attachment for the AT1-R and has an extremely long duration of half-life among the ARBs ^[60]. Furthermore, Telmisartan's lipophilicity and preference for the liver make it ideal for liver signs ^[60]. The angiotensin-converting enzyme (ACE) 2/Ang (1-7) process was investigated just like a substitute to that of the RAS ^[61]. The stimulation of Mas, the G-protein-coupled Ang (1-7) receptor, induces vasodilation and antifibrotic signaling pathways in heart myocytes ^[62]. In this process, ACE2 changes Ang II to Ang (1–7) and Ang I to Ang (1–9) ^[63]. Telmisartan has been shown to improve hepatic fibrosis markers in various destructive contexts ^[64].

Telmisartan's therapeutic impact on hepatocytes can be linked to its anti-oxidative properties ^[51]. likewise, telmisartan decreased liver fibrosis that was triggered in rats by diet with methioninedeficient and choline-deficient ^[65]. Telmisartan affects the production of PPAR-y target genes that regulate the metabolism of carbohydrates and lipids. It has been demonstrated to decrease levels of glucose, insulin, and triglyceride in animals fed a high-fat, high-carbohydrate diet ^[49]. PPAR-y improves insulin sensitivity, and high-density and reduces levels lipoprotein levels, of inflammation, oxidative stress, cell migration and proliferation, and fatty acid and triglyceride. However, it does not cause accumulation of fluid like full agonists of PPAR- γ , such as pioglitazone or rosiglitazone^[51] as shown in *Figure 6*.

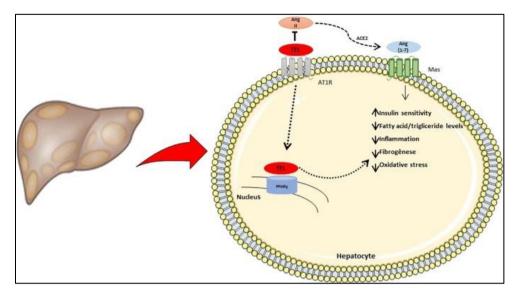


Figure 6. The liver-protecting actions of telmisartan in liver diseases are shown ^[66].

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REFERENCE

- Lee Y A, Wallace M C, and Friedman S L. Pathobiology of liver fibrosis: a translational success story. Gut. 2015; 64(5): p. 830-841.
- Aydın M M and Akçalı K C. Liver fibrosis. The Turkish Journal of Gastroenterology. 2018; 29(1): p. 14.
- Wang F S, Fan J G, Zhang Z, Gao B, and Wang H Y. The global burden of liver disease: the major impact of China. Hepatology. 2014; 60(6): p. 2099-2108.
- 4. *Park B-J, Lee Y-J, and Lee H-R.* Chronic liver inflammation: clinical implications beyond alcoholic liver disease. World journal of gastroenterology. 2014; 20(9): p. 2168-2199.
- Osna N A, Donohue Jr T M, and Kharbanda K K. Alcoholic Liver Disease: Pathogenesis and Current Management. Alcohol Research: Current Reviews. 2017; 38(2): p. 147-159.

- Pellicoro A, Ramachandran P, Iredale J P, and Fallowfield J A. Liver fibrosis and repair: immune regulation of wound healing in a solid organ. Nature Reviews Immunology. 2014; 14(3): p. 181-194.
- Bataller R and Brenner D A. Liver fibrosis. The Journal of Clinical Investigation. 2005; 115(2): p. 209-218.
- 8. *Kisseleva T and Brenner D A.* Mechanisms of fibrogenesis. Experimental biology and medicine. 2008; 233(2): p. 109-122.
- Grigorescu M. Noninvasive biochemical markers of liver fibrosis. Journal of gastrointestinal and liver diseases: JGLD. 2006; 15(2): p. 149-159.
- Carpino G, Morini S, Corradini S G, Franchitto A, Merli M, Siciliano M, Gentili F, Muda A O, Berloco P, and Rossi M. Alpha-SMA expression in hepatic stellate cells and quantitative analysis of hepatic fibrosis in cirrhosis and in recurrent chronic hepatitis after liver transplantation. Digestive and Liver Disease. 2005; 37(5): p. 349-356.
- Ma X. Liver fibrogenesis in non-alcoholic steatohepatitis. Frontiers in physiology. 2012; 3: p. 248.

- Seki E and Brenner D A. Recent advancement of molecular mechanisms of liver fibrosis. Journal of Hepato-Biliary-Pancreatic Sciences. 2015; 22(7): p. 512-518.
- 13. *Kisseleva T.* The origin of fibrogenic myofibroblasts in fibrotic liver. Hepatology. 2017; 65(3): p. 1039-1043.
- Iwaisako K, Jiang C, Zhang M, Cong M, Moore-Morris T J, Park T J, Liu X, Xu J, Wang P, and Paik Y-H. Origin of myofibroblasts in the fibrotic liver in mice. Proceedings of the National Academy of Sciences. 2014; 111(32): p. E3297-E3305.
- Karin D, Koyama Y, Brenner D, and Kisseleva T. The characteristics of activated portal fibroblasts/myofibroblasts in liver fibrosis. Differentiation. 2016; 92(3): p. 84-92.
- El Agha E, Kramann R, Schneider R K, Li X, Seeger W, Humphreys B D, and Bellusci S. Mesenchymal stem cells in fibrotic disease. Cell stem cell. 2017; 21(2): p. 166-177.
- Vickers N J. Animal communication: when I'm calling you, will you answer too? Current biology. 2017; 27(14): p. R713-R715.
- Khomich O, Ivanov A V, and Bartosch B. Metabolic hallmarks of hepatic stellate cells in liver fibrosis. Cells. 2020; 9(1): p. 24.
- Senoo H, Mezaki Y, and Fujiwara M. The stellate cell system (vitamin A-storing cell system). Anatomical Science International. 2017; 92(4): p. 387-455.
- 20. Peng W, Cheng S, Bao Z, Wang Y, Zhou W, Wang J, Yang Q, Chen C, and Wang W. Advances in the research of nano drug delivery system for targeted treatment of liver fibrosis. Biomedicine & Pharmacotherapy. 2021; 137: p. 111342.
- 21. Nallagangula K S, Nagaraj S K, Venkataswamy L, and Chandrappa M.

Liver fibrosis: a compilation on the biomarkers status and their significance during disease progression. Future Sci OA. 2018; 4(1): p. Fso250.

- Bedossa P, Dargère D, and Paradis V. Sampling variability of liver fibrosis in chronic hepatitis C. Hepatology. 2003; 38(6): p. 1449-1457.
- 23. Braticevici C F, Papacocea R, Tribus L, and Badarau A. Can We Replace Liver Biopsy with Non-Invasive Procedures? Liver biopsy. 2011: p. 225-240.
- Goodman Z D. Grading and staging systems for inflammation and fibrosis in chronic liver diseases. Journal of Hepatology. 2007; 47(4): p. 598-607.
- Pinzani M, Rombouts K, and Colagrande S. Fibrosis in chronic liver diseases: diagnosis and management. Journal of Hepatology. 2005; 42(1): p. S22-S36.
- Gressner A M, Gao C-F, and Gressner O
 A. Non-invasive biomarkers for monitoring the fibrogenic process in liver: a short survey. World Journal of Gastroenterology: WJG. 2009; 15(20): p. 2433.
- 27. Dufour D R, Lott J A, Nolte F S, Gretch D R, Koff R S, and Seeff L B. Diagnosis and monitoring of hepatic injury. II. Recommendations for use of laboratory tests in screening, diagnosis, and chemistry. 2000; monitoring. Clinical 46(12): p. 2050-2068.
- 28. *Kwo P Y, Cohen S M, and Lim J K.* ACG clinical guideline: evaluation of abnormal liver chemistries. Official journal of the American College of Gastroenterology| ACG. 2017; 112(1): p. 18-35.
- Afdhal N H and Nunes D. Evaluation of liver fibrosis: a concise review. Official journal of the American College of Gastroenterology ACG. 2004; 99(6): p. 1160-1174.

- 30. *Ebrahimi H, Naderian M, and Sohrabpour A A.* New concepts on reversibility and targeting of liver fibrosis; a review article. Middle East Journal of digestive diseases. 2018; 10(3): p. 133.
- Koyama Y, Xu J, Liu X, and Brenner D A. New developments on the treatment of liver fibrosis. Digestive diseases. 2016; 34(5): p. 589-596.
- 32. Ma R, Chen J, Liang Y, Lin S, Zhu L, Liang X, and Cai X. Sorafenib: A potential therapeutic drug for hepatic fibrosis and its outcomes. Biomedicine & Pharmacotherapy. 2017; 88: p. 459-468.
- Anderson P H, Turner A G, and Morris H
 A. Vitamin D actions to regulate calcium and skeletal homeostasis. Clinical Biochemistry. 2012; 45(12): p. 880-886.
- Hossein-nezhad A and Holick M F. Vitamin D for health: a global perspective. in Mayo Clinic proceedings. 2013. Elsevier.
- 35. Noreña J A, Niño C D, Gallego S, Builes-Barrera C A, Castro D C, Román-González A, and Jimenez C. Calcitriol-mediated hypercalcemia secondary to granulomatous disease caused by soft-tissue filler injection: a case report. Clinical Cases in Mineral and Bone Metabolism. 2017; 14(3): p. 340.
- 36. Gascon-Barré M, Demers C, Mirshahi A, Néron S, Zalzal S, and Nanci A. The normal liver harbors the vitamin D nuclear receptor in nonparenchymal and biliary epithelial cells. Hepatology. 2003; 37(5): p. 1034-1042.
- 37. Udomsinprasert W and Jittikoon J.
 Vitamin D and liver fibrosis: Molecular mechanisms and clinical studies.
 Biomedicine & pharmacotherapy. 2019; 109: p. 1351-1360.
- 38. Garland C F, Garland F C, Gorham E D, Lipkin M, Newmark H, Mohr S B, and Holick M F. The role of vitamin D in cancer

prevention. American journal of public health. 2006; 96(2): p. 252-261.

- 39. Abramovitch S, Dahan-Bachar L, Sharvit E, Weisman Y, Tov A B, Brazowski E, and Reif S. Vitamin D inhibits proliferation and profibrotic marker expression in hepatic stellate cells and decreases thioacetamideinduced liver fibrosis in rats. Gut. 2011; 60(12): p. 1728-1737.
- 40. Neeman R, Abramovitch S, Sharvit E, Elad-Sfadia G, Haklai R, Kloog Y, and Reif S. Vitamin D and S-farnesylthiosalicylic acid have a synergistic effect on hepatic stellate cells proliferation. Digestive diseases and sciences. 2014; 59: p. 2462-2469.
- 41. *Artaza J N and Norris K C.* Vitamin D reduces the expression of collagen and key profibrotic factors by inducing an antifibrotic phenotype in mesenchymal multipotent cells. Journal of Endocrinology. 2009; 200(2): p. 207.
- 42. Liu Y, Liu H, Meyer C, Li J, Nadalin S, Königsrainer A, Weng H, Dooley S, and Ten Dijke P. Transforming growth factor-β (TGF-β)-mediated connective tissue growth factor (CTGF) expression in hepatic stellate cells requires Stat3 signaling activation. Journal of Biological Chemistry. 2013; 288(42): p. 30708-30719.
- 43. Wang L, Yuan T, Du G, Zhao Q, Ma L, and **J**. The of Zhu impact 1, 25dihydroxyvitamin D3 on the expression of connective tissue growth factor and transforming growth factor- β 1 in the myocardium of rats with diabetes. Diabetes Research and Clinical Practice. 2014; 104(2): p. 226-233.
- Honsawek S, Udomsinprasert W, Chirathaworn C, Anomasiri W, Vejchapipat P, and Poovorawan Y. Correlation of connective tissue growth factor with liver stiffness measured by

transient elastography in biliary atresia. Hepatology Research. 2013; 43(7): p. 795-800.

- 45. Beilfuss A, Sowa J-P, Sydor S, Beste M, Bechmann L P, Schlattjan M, Syn W-K, Wedemeyer I, Mathé Z, and Jochum C. Vitamin D counteracts fibrogenic TGF-β signaling in human hepatic stellate cells both receptor-dependently and independently. Gut. 2015; 64(5): p. 791-799.
- 46. Fisher L and Fisher A. Vitamin D and parathyroid hormone in outpatients with noncholestatic chronic liver disease. Clinical gastroenterology and Hepatology. 2007; 5(4): p. 513-520.
- 47. Arteh J, Narra S, and Nair S. Prevalence of vitamin D deficiency in chronic liver disease. Digestive diseases and sciences. 2010; 55(9): p. 2624-2628.
- 48. Zittermann A, Iodice S, Pilz S, Grant W B, Bagnardi V, and Gandini S. Vitamin D deficiency and mortality risk in the general population: a meta-analysis of prospective cohort studies. The American journal of clinical nutrition. 2012; 95(1): p. 91-100.
- 49. Benson S C, Pershadsingh H A, Ho C I, Chittiboyina A, Desai P, Pravenec M, Qi N, Wang J, Avery M A, and Kurtz T W. Identification of telmisartan as a unique angiotensin II receptor antagonist with selective PPARγ-modulating activity. Hypertension. 2004; 43(5): p. 993-1002.
- 50. Yokohama S, Tokusashi Y, Nakamura K, Tamaki Y, Okamoto S, Okada M, Aso K, Hasegawa T, Aoshima M, and Miyokawa N. Inhibitory effect of angiotensin II receptor antagonist on hepatic stellate cell activation in non-alcoholic steatohepatitis. World journal of gastroenterology: WJG. 2006; 12(2): p. 322.
- 51. de Macêdo S M, Guimarães T A, Feltenberger J D, and Santos S H S. The

role of renin-angiotensin system modulation on treatment and prevention of liver diseases. Peptides. 2014; 62: p. 189-196.

- 52. Halici Z, Bilen H, Albayrak F, Uyanik A, Cetinkaya R, Suleyman H, Keles O N, and Unal B. Does telmisartan prevent hepatic fibrosis in rats with alloxan-induced diabetes? European journal of pharmacology. 2009; 614(1-3): p. 146-152.
- 53. Marui N, Offermann M K, Swerlick R, Kunsch C, Rosen C A, Ahmad M, Alexander R W, and Medford R M. Vascular cell adhesion molecule-1 (VCAM-1) gene transcription and expression are regulated through an antioxidant-sensitive mechanism in human vascular endothelial cells. The Journal of Clinical Investigation. 1993; 92(4): p. 1866-1874.
- 54. *Yi E-t, Liu R-x, Wen Y, and Yin C-h.* Telmisartan attenuates hepatic fibrosis in bile duct-ligated rats. Acta Pharmacologica Sinica. 2012; 33(12): p. 1518-1524.
- 55. Cynis H, Kehlen A, Haegele M, Hoffmann T, Heiser U, Fujii M, Shibazaki Y, Yoneyama H, Schilling S, and Demuth H U. Inhibition of Glutaminyl Cyclases alleviates CCL 2-mediated inflammation of non-alcoholic fatty liver disease in mice. International journal of experimental pathology. 2013; 94(3): p. 217-225.
- 56. Bakheit A H, Abd-Elgalil A A, Mustafa B, Haque A, and Wani T A. Telmisartan. Profiles of drug substances, excipients and related methodology. 2015; 40: p. 371-429.
- 57. Georgescu E F, Ionescu R, Niculescu M, Mogoanta L, and Vancica L. Angiotensinreceptor blockers as therapy for mild-tomoderate hypertension-associated nonalcoholic steatohepatitis. World journal of gastroenterology: WJG. 2009; 15(8): p. 942.
- 58. Soler M J, Ye M, Wysocki J, William J,
 Lloveras J, and Batlle D. Localization of
 ACE2 in the renal vasculature:

amplification by angiotensin II type 1 receptor blockade using telmisartan. American Journal of Physiology-Renal Physiology. 2009; 296(2): p. F398-F405.

- 59. Bataller R, Gäbele E, Parsons C J, Morris T, Yang L, Schoonhoven R, Brenner D A, and Rippe R A. Systemic infusion of angiotensin II exacerbates liver fibrosis in bile duct–ligated rats. Hepatology. 2005; 41(5): p. 1046-1055.
- Schuppan D, Gorrell M D, Klein T, Mark M, and Afdhal N H. The challenge of developing novel pharmacological therapies for non-alcoholic steatohepatitis. Liver International. 2010; 30(6): p. 795-808.
- 61. *Iwai M and Horiuchi M.* Devil and angel in the renin-angiotensin system: ACE– angiotensin II–AT1 receptor axis vs. ACE2– angiotensin-(1–7)–Mas receptor axis. Hypertension Research. 2009; 32(7): p. 533-536.
- 62. Tallant E A, Ferrario C M, and Gallagher P E. Angiotensin-(1–7) inhibits growth of cardiac myocytes through activation of the mas receptor. American Journal of Physiology- Heart and circulatory physiology. 2005; 289(4): p. H1560-H1566.

 Li N, Zimpelmann J, Cheng K, Wilkins J A, and Burns K D. The role of angiotensin converting enzyme 2 in the generation of angiotensin 1–7 by rat proximal tubules. American Journal of Physiology-Renal Physiology. 2005; 288(2): p. F353-F362.

pISSN: 2636-4093, eISSN: 2636-4107

- 64. *Alberti K G M M, Zimmet P, and Shaw J.* Metabolic syndrome—a new world-wide definition. A consensus statement from the international diabetes federation. Diabetic medicine. 2006; 23(5): p. 469-480.
- 65. *Tamaki Y, Nakade Y, Yamauchi T, Makino Y, Yokohama S, Okada M, Aso K, Kanamori H, Ohashi T, and Sato K.* Angiotensin II type 1 receptor antagonist prevents hepatic carcinoma in rats with nonalcoholic steatohepatitis. Journal of Gastroenterology. 2013; 48: p. 491-503.
- 66. Borém L M A, Neto J F R, Brandi I V, Lelis D F, and Santos S H S. The role of the angiotensin II type I receptor blocker telmisartan in the treatment of non-alcoholic fatty liver disease: a brief review. Hypertension Research. 2018; 41(6): p. 394-405.